

INVESTIGATION OF ANTIMICROBIAL ACTIVITIES OF CAULERPA MEXICANA AND UDOTEA LONGLIFERA AGAINST SURFACE ASSOCIATED STREPTOCOCCUS KUTZSCHKEI

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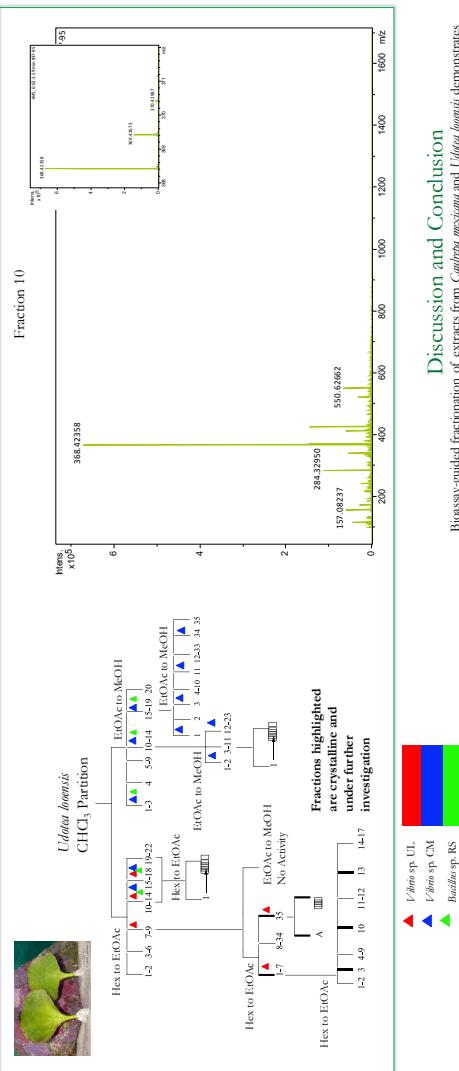
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Abstract

In a broad survey of extant marine algae from the Florida Keys against an extensive panel of environmental bacteria, from the green alga *T. sp.* to one strain of *Raffelia* sp. To continue the significant activity against two strains of *T. sp.*, the crude extract was subjected to investigation of the antibacterial activity of the culture collected by stockpiling (Hord Day, Hord Keys) biomass/gel fractionation [1]. It was immediately apparent at 20 degrees Celsius and 24 hours followed by a depth of 1.5 meters in July 2012 and 2013, multiple active fractions were isolated by NMR on a *TOUGH* 400 MHz NMR. Extracts and column fractions were assayed against three strains of bacteria maintained in A medium. *Vibrio fischeri*, *Vibrio sp.* and *Salmonella enterica* serotype *Altona* were used as model systems. Initial and final reading were measured using a Biotech plate reader (Biorad) at 25mL and 247 hours sample dependent. Cetazoline fractions were analyzed on a Biotech Biorad Vario KL 1000 with 5 mm protein (TCI/ISN/TM) Cetazoline Probe. Mass spectra were obtained on an ESI (ultra high resolution QTOF QM/MS Maxx 4G) in 1 in 1000 dilution of 91c15 mixture of acetone/tic to water of a working stock, 1mg/ml solution.

Results



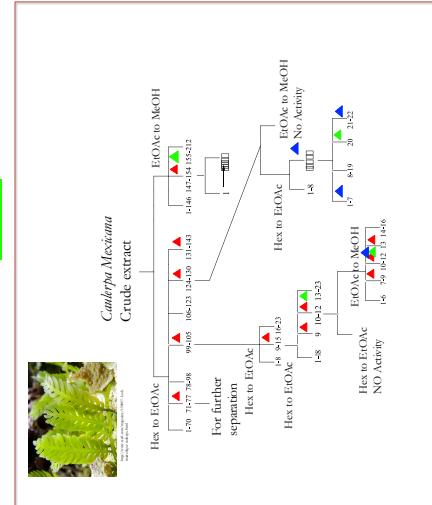
Discussion and Conclusion

These data suggest that *C. mucicola* produces a complex mixture of metabolites that inhibit the growth of *Vibrio* sp. UL, *Vibrio* sp. CM and *Bacillus* sp. RS. Acting is mainly seen in the nonpolar fractions of *C. mucicola* against *Vibrio* sp. UL. We also see activity against *Vibrio* sp. UL in the polar fractions of *C. mucicola*, while that more polar fractions contain metabolites that inhibit the growth of *Vibrio* sp. CM and *Bacillus* sp. RS. Fractionation of *C. mucicola* yields one fraction with crystals. The MS data shows that this molecule could be potentially be a novel antimicrobial compound.

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The authors wish to thank students Mitchell Jef, Kyle Nevers, Ian B. Newby, Leah Meador, and Michaela Deane for collecting hair samples and their assistance with experiments for this study and the two anonymous reviewers. We thank Brian Murphy (the Kappa Delta chapter at the University of Alberta) for his help with the statistical analysis and Dr. David P. Miettinen, a former student at the University of Alberta, for his help with the manuscript preparation. We also thank Dr. Michael J. Hickey (the Kappa Delta chapter at the University of Alberta) for his help with the manuscript preparation. This research was funded by the College of Pharmacy CSE – thank you! This research is funded by the College of Pharmacy CSE. This project was conducted as part of the Capstone Research Project and Summer Student Research Program at the College of Pharmacy CSE.

Dose-response



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Metabolism
Cadmus meianus and *Udotea kozai* were collected by snorkeling from the Florida Bay in the fall of 2012 and immediately frozen at -3 °C. Larvae were collected by snorkeling from the Florida Bay in 2012 and 2013, and were immediately frozen at -3 °C, and transported to CSU, College of Marine and Coastal Sciences, and stored at -80 °C. All larvae were extracted in 10 mM Tris-HCl, pH 7.4, over 2 hours and dried. *C. meianus* was partitioned between H₂O and D₂O. *C. meianus* was subjected to repeated Si gel chromatography using a medium pressure liquid chromatograph apparatus Biogel P-600. Once to yield multiple active fractions. *U. kozai* was subjected to repeated Si gel chromatography apparatus Biogel P-600. Once to yield multiple active fractions. *U. kozai* was partitioned between H₂O and D₂O. The ED₅₀ for each fraction was further analyzed by HPLC on a Waters Acclaim column (2.1 mm x 150 mm) at 35 °C. CH₃OH/H₂O/MOM (10:90 v/v/v) was used as the mobile phase. The controls were streptomycin, kanamycin, tetracycline, and DMSO. The initial and final reading on the spectrophotometer were read every 24 hours, depending on the time of collection. The controls were streptomycin, kanamycin, tetracycline, and DMSO. The initial and final reading on the spectrophotometer were read every 24 hours, depending on the time of collection. *U. kozai* and *Bathyporeia* were assayed against three environmental strains of bacteria. *Enterobacter cloacae*, *Escherichia coli*, and *Salmonella enterica* serotype *Infantis* were isolated from the surface of *C. meianus*. Vibrioplymnae was isolated from the surface of *C. meianus*. *Vibrio* sp. was isolated from the surface of *U. kozai*. *Escherichia coli* and *Salmonella enterica* serotype *Infantis* were isolated from the surface of *Bathyporeia*. All liquid media, solid media, and diluent of DMSO-excuted were added to each well of a 96-well plate. The controls were streptomycin, kanamycin, tetracycline, and DMSO. The initial and final reading on the spectrophotometer were read every 24 hours, depending on the time of collection. The controls were streptomycin, kanamycin, tetracycline, and DMSO. Mass spectra were obtained on a LSI ultra high resolution QTOF LC/MS/MS (MicrMass Q-ToF Premier). A 1 μM solution of actinomycete extract was prepared in 100% methanol. One microliter of the actinomycete extract was added to 990 μL of water of a weight: stock (1:100) mixture, one solution.